

Using asreml and asremlPlus for the Ladybird example from Welham et al. (2014)

Chris Brien

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Introduction

This vignette shows how to use `asremlPlus` (Brien, 2025a), and `dae` (Brien, 2024b), for exploring and presenting predictions (estimated marginal means: EMMs) from a linear mixed model analysis, the predictions having been produced using `asreml` (Butler et al., 2023). Here, `asremlPlus`, `dae` and `asreml` are packages for the R Statistical Computing environment (R Core Team, 2025).

The context is a three-factor factorial experiment on ladybirds (Welham et al., 2014, Example 8.2) that aims to answer the question “Will ladybirds transfer fungus to aphids on plants?” The experiment consists of 2 runs of 36 containers, each with a plant and aphids. There are three factors that results in 12 treatments: Host plant (beans, trefoil), infected Cadavers (5, 10, 20), Ladybird (-, +). These are randomized to the containers within a run so that each is replicated 3 times within a run. The response to be analysed is the logit of the proportion of live aphids that were infected.

Initialize

```
library(knitr)
opts_chunk$set("tidy" = FALSE, comment = NA)
suppressMessages(library(asreml))
```

```
## Offline License checked out Tue Jun 10 18:07:40 2025
```

```
packageVersion("asreml")
```

```
## [1] '4.2.0.370'
```

```
suppressMessages(library(asremlPlus))
packageVersion("asremlPlus")
```

```
## [1] '4.4.49'
```

```
suppressMessages(library("dae"))
packageVersion("dae")
```

```
## [1] '3.2.30'
```

```
options(width = 95, show.signif.stars = FALSE)
```

Get data available in asremlPlus

```
data("Ladybird.dat")
```

Do an ANOVA of logits

```
Ladybird.aov <- aov(logitP ~ Host*Cadavers*Ladybird + Error(Run/Plant),
                    data=Ladybird.dat)
summary(Ladybird.aov)
```

```
Error: Run
      Df Sum Sq Mean Sq F value Pr(>F)
Residuals 1 0.06766 0.06766

Error: Run:Plant
      Df Sum Sq Mean Sq F value Pr(>F)
Host      1 13.599  13.599  59.172 1.82e-10
Cadavers  2 17.027   8.514  37.044 3.78e-11
Ladybird  1 11.091  11.091  48.257 3.33e-09
Host:Cadavers  2  0.308   0.154   0.670  0.5158
Host:Ladybird  1  0.228   0.228   0.992  0.3234
Cadavers:Ladybird  2  1.735   0.867   3.774  0.0287
Host:Cadavers:Ladybird  2  0.200   0.100   0.435  0.6493
Residuals      59 13.560   0.230
```

The anova table gives the F-tests for the three-factor effects and interactions. Note the **Residuals** **Mean Sq** value for Run:Plant of 0.230. Also, it is clear that the Run component is negative, given that the **Residuals** **Mean Sq** value for Run is less than that for Run:Plant; it is $(0.06766 - 0.230) / 36$. From the table it is seen that the only significant interaction is Cadavers:Ladybird and that the Host main effect is significant.

Use asreml to analyse the logits

Mixed model analysis of logits

```
m <- asreml(logitP ~ Host*Cadavers*Ladybird,
            random = ~ Run,
            residual = ~ Run:Plant,
            data = Ladybird.dat)
```

```
ASReml Version 4.2 10/06/2025 18:07:41
      LogLik      Sigma2      DF      wall
1      3.071301    0.2262391     60    18:07:41 ( 1 restrained)
```

```

2      3.642605      0.2269125      60      18:07:41      ( 1 restrained)
3      3.712498      0.2271056      60      18:07:41      ( 1 restrained)
4      3.717213      0.2271206      60      18:07:41      ( 1 restrained)
5      3.717512      0.2271216      60      18:07:41      ( 1 restrained)

```

```
summary(m)$varcomp
```

```

              component  std.error  z.ratio bound %ch
Run           2.298309e-08         NA      NA      B  NA
Run:Plant!R    2.271216e-01  0.04156985  5.463612      P   0

```

As expected the Run component is bound (B) at approximately zero. This results in a change in the estimate of the residual variance to 0.227. To allow for a negative estimate we will unconstrain the Run component. As Littell et al. (2006, p.150) say

if you do not set the negative variance component estimate to zero, but allow it to remain negative, you get better control over Type I error and, for cases of negative wholeplot error variance estimates, greater power. Therefore, this is the recommended procedure.

Unconstrain Reps to make the analysis equivalent to ANOVA

```
m <- setvarianceterms(m$call, terms = "Run", bounds = "U")
```

```

ASReml Version 4.2 10/06/2025 18:07:41
      LogLik      Sigma2      DF      wall
1      3.071301      0.2262391      60      18:07:41      ( 1 restrained)
2      3.642605      0.2269125      60      18:07:41      ( 1 restrained)
3      3.802834      0.2274541      60      18:07:41      ( 1 restrained)
4      3.839487      0.2334176      60      18:07:41
5      3.955683      0.2309710      60      18:07:41
6      3.973334      0.2300353      60      18:07:41
7      3.974051      0.2298346      60      18:07:41

```

```
summary(m)$varcomp
```

```

              component  std.error  z.ratio bound %ch
Run           -0.004504789  0.002896281 -1.555370      U 0.1
Run:Plant!R    0.229834648  0.042316936  5.431269      P 0.0

```

Now the Run component is negative and the Run:Plant variance estimate is now equal to that for the Residuals Mean Sq for Run:Plant from the anova table.

Set up an asrtests object

```

current.asrt <- as.asrtests(m)
print(current.asrt, which = "pseudoanova")

```

```
#### Pseudo-anova table for fixed terms
```

Wald tests for fixed effects.

Response: logitP

	Df	denDF	F.inc	Pr
(Intercept)	1	1	1550.00	0.0162
Host	1	59	59.17	0.0000
Cadavers	2	59	37.04	0.0000
Ladybird	1	59	48.26	0.0000
Host:Cadavers	2	59	0.67	0.5158
Host:Ladybird	1	59	0.99	0.3234
Cadavers:Ladybird	2	59	3.77	0.0287
Host:Cadavers:Ladybird	2	59	0.44	0.6493

The `asrtests` object contains a `wald.tab` component which can be printed by specifying that the `pseudoanova` is printed. The F -values for the fixed terms in this table are the same as those in the anova table.

Obtain the marginality matrix for the fixed terms

The `pstructure` function from the `dae` package (Brien, 2024b) produce the marginality matrix for a formula as a side effect and we take advantage of that to obtain the matrix required here.

```
Ladybird.pstr <- pstructure(formula = ~ Host*Cadavers*Ladybird,
                             data = Ladybird.dat)
HCL.marg <- marginality(Ladybird.pstr)
print(HCL.marg)
```

	Host	Cadavers	Host:Cadavers	Ladybird	Host:Ladybird	Cadavers:Ladybird
Host	1	0	1	0	1	0
Cadavers	0	1	1	0	0	1
Host:Cadavers	0	0	1	0	0	0
Ladybird	0	0	0	1	1	1
Host:Ladybird	0	0	0	0	1	0
Cadavers:Ladybird	0	0	0	0	0	1
Host:Cadavers:Ladybird	0	0	0	0	0	0

	Host:Cadavers:Ladybird
Host	1
Cadavers	1
Host:Cadavers	1
Ladybird	1
Host:Ladybird	1
Cadavers:Ladybird	1
Host:Cadavers:Ladybird	1

This marginality matrix is interpreted by taking a row term and noting that it is marginal to any column term with a one in this row.

Choose marginality-compliant model

```
chosen <- chooseModel(current.asrt, terms.marginality = HCL.marg)
current.asrt <- chosen$asrtests.obj
print(current.asrt, which = "test", omit.columns = c("AIC", "BIC"))
```

Sequence of model investigations for logitP

	terms	DF	denDF	p	action
1	Host:Cadavers:Ladybird	2	59	0.6493	Nonsignificant
2	Cadavers:Ladybird	2	59	0.0287	Significant
3	Host:Ladybird	1	59	0.3234	Nonsignificant
4	Host:Cadavers	2	59	0.5158	Nonsignificant
5	Host	1	59	0.0000	Significant

```
(chosen$sig.terms)
```

```
[[1]]
```

```
[1] "Cadavers:Ladybird"
```

```
[[2]]
```

```
[1] "Host"
```

The `chooseModel` function produces a list with components `sig.terms`, a list with the terms in the marginality-compliant model, and `asrtests.obj`, the `asrtests` object resulting from the model selection. In particular, the `asrtests` object contains a `test.summary` that details the tests performed in choosing the model; the AIC and BIC columns are omitted from `test.summary` because their inclusion has not been requested. Note that `chooseModel` does not test the main effects for Cadavers or Ladybird, because these are marginal to the significant two-factor interaction Cadavers:Ladybird.

Form formula for selected model

```
chosen.mod <- paste(unlist(chosen$sig.terms), collapse = " + ")
(chosen.mod <- as.formula(paste("~", chosen.mod)))
```

```
~Cadavers:Ladybird + Host
```

Obtain predictions under the chosen model and form an alldiffs object

```
diffs <- predictPlus(current.asrt$asreml.obj,
  classify = "Host:Ladybird:Cadavers",
  linear.transformation = ~Cadavers:Ladybird + Host,
  wald.tab = current.asrt$wald.tab,
  error.intervals = "halfLeast",
  LSDtype = "factor.combination", LSDby = "Host",
  tables = "predictions")
```

```
Joining with 'by = join_by(fac.comb)'  
Joining with 'by = join_by(Host)'
```

Predictions for logitP transform(s) from Host:Ladybird:Cadavers

Notes:

The original predictions, obtained as described below, have been linearly transformed to form estimated marginal means.

- The predictions are obtained by averaging across the hypertable calculated from model terms constructed solely from factors in the averaging and classify sets.
- Use 'average' to move ignored factors into the averaging set.
- The ignored set: Run

	Host	Ladybird	Cadavers	predicted.value	standard.error	upper.halfLeastSignificant.limit
1	bean	-	5	-1.6038338	0.1417454	-1.4080222
2	bean	-	10	-1.1454308	0.1417454	-0.9496192
3	bean	-	20	-0.7448097	0.1417454	-0.5489981
4	bean	+	5	-1.0195475	0.1417454	-0.8237359
5	bean	+	10	-0.5983440	0.1417454	-0.4025323
6	bean	+	20	0.4786704	0.1417454	0.6744820
7	trefoil	-	5	-2.4730339	0.1417454	-2.2772223
8	trefoil	-	10	-2.0146309	0.1417454	-1.8188193
9	trefoil	-	20	-1.6140098	0.1417454	-1.4181982
10	trefoil	+	5	-1.8887476	0.1417454	-1.6929360
11	trefoil	+	10	-1.4675441	0.1417454	-1.2717325
12	trefoil	+	20	-0.3905297	0.1417454	-0.1947181

	lower.halfLeastSignificant.limit	est.status
1	-1.7996454	Estimable
2	-1.3412425	Estimable
3	-0.9406214	Estimable
4	-1.2153592	Estimable
5	-0.7941556	Estimable
6	0.2828588	Estimable
7	-2.6688455	Estimable
8	-2.2104426	Estimable
9	-1.8098215	Estimable
10	-2.0845593	Estimable
11	-1.6633557	Estimable
12	-0.5863414	Estimable

LSD values

minimum LSD = 0.3916233 0.3916233

mean LSD = 0.3916233 0.3916233

maximum LSD = 0.3916233 0.3916233

(sed range / mean sed = 1.28e-15 1.13e-15)

Setting the `terms` argument to `Host:Ladybird:Cadavers` requests predictions for all combinations of the three factors and the `linear.transformation` argument is used to obtain estimated marginal means (EMMs) that conform to the chosen model. The `wald.tab` is supplied so that it can be used to get the degrees of freedom for the *t*-value to be used in calculating the LSD; the degrees of freedom to the source for the `terms` argument will be used. The `error.intervals` argument has been set to `"halfLeast"`, the `LSDtype` argument to `"factor.combination"` and the `LSDby` argument to `"Host"` so that the average LSD will be calculated for each Host. This necessary because, under the chosen model, the LSDs differ between Hosts. It results in `lower.halfLeastSignificant.limit` and `upper.halfLeastSignificant.limit` being added to the `predictions` component of the `alldiffs` object.

Or, calculate predictions to check first and then transform to conform to chosen model

```
diffs.full <- predictPlus(current.asrt$asreml.obj,
                          classify = "Host:Ladybird:Cadavers",
                          wald.tab = current.asrt$wald.tab,
                          tables = "none", Vmatrix = TRUE)

diffs <- linTransform(diffs.full, linear.transformation = ~Cadavers:Ladybird + Host,
                      wald.tab = current.asrt$wald.tab,
                      error.intervals = "halfLeast",
                      LSDtype = "factor.combination", LSDby = "Host",
                      tables = "predictions")
```

Joining with `'by = join_by(fac.comb)'`

Joining with `'by = join_by(Host)'`

Predictions for logitP transform(s) from Host:Ladybird:Cadavers

Notes:

The original predictions, obtained as described below, have been linearly transformed to form estimated marginal means.

- The predictions are obtained by averaging across the hypertable calculated from model terms constructed solely from factors in the averaging and classify sets.
- Use `'average'` to move ignored factors into the averaging set.
- The ignored set: Run

	Host	Ladybird	Cadavers	predicted.value	standard.error	upper.halfLeastSignificant.limit
1	bean	-	5	-1.6038338	0.1417454	-1.4080222
2	bean	-	10	-1.1454308	0.1417454	-0.9496192
3	bean	-	20	-0.7448097	0.1417454	-0.5489981
4	bean	+	5	-1.0195475	0.1417454	-0.8237359
5	bean	+	10	-0.5983440	0.1417454	-0.4025323
6	bean	+	20	0.4786704	0.1417454	0.6744820
7	trefoil	-	5	-2.4730339	0.1417454	-2.2772223
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10	trefoil	+	5	-1.8887476	0.1417454	-1.6929360
11	trefoil	+	10	-1.4675441	0.1417454	-1.2717325

12	trefoil	+	20	-0.3905297	0.1417454	-0.1947181
	lower.halfLeastSignificant.limit est.status					
1				-1.7996454	Estimable	
2				-1.3412425	Estimable	
3				-0.9406214	Estimable	
4				-1.2153592	Estimable	
5				-0.7941556	Estimable	
6				0.2828588	Estimable	
7				-2.6688455	Estimable	
8				-2.2104426	Estimable	
9				-1.8098215	Estimable	
10				-2.0845593	Estimable	
11				-1.6633557	Estimable	
12				-0.5863414	Estimable	

LSD values

minimum LSD = 0.3916233 0.3916233

mean LSD = 0.3916233 0.3916233

maximum LSD = 0.3916233 0.3916233

(sed range / mean sed = 1.28e-15 1.13e-15)

The above LSD values can only be used to compare pairs of EMMs for the same Host.

Explore the LSDs

To investigate the errors that would result from using the overall LSDs as opposed to the LSDs computed for each Host, the `exploreLSDs`, `pickLSDstatistics` and `plotLSDerrors` functions are used, firstly with the default value of "overall" for `LSDtype` and finally with the `LSDtype` set to "factor.combination" and `LSDby` to "Host".

The `exploreLSDs` function produces tables of statistics for the LSDs computed for the settings of the `LSDtype` and `LSDby`; the settings of these arguments does not have to match those used in producing the `alldiffs` object. For `LSDtype` set to "overall", a single LSD statistic is computed that is based on the standard errors of all pairwise differences. To ascertain the errors that arise from using this LSD value for determining, for all pairwise comparisons, whether a comparison is significant, `exploreLSDs` compares the results using the LSD value with the p -values in the `p.differences` component of the `alldiffs` object. For `LSDtype` set to "factor.combination" and `LSDby` to "Host", the LSD statistics are calculated from standard errors of the pairwise differences for each Host. Examination of the `sed` component of `diffs` reveals that there are only three different values for the standard errors of pairwise differences and, hence, only three unique values for the LSD. Of the statistics shown, there is no single value that will not result in errors.

Here are the tables of the numbers of false positive and negative error in using the values of the various LSD statistics for determining the significance of the 66 pairwise comparisons of the 12 predictions.

```
exploreLSDs(diffs, LSDtype = "overall")
```

```
#### Statistics calculated from LSD values
```


	c	min	quant10	quant25	mean	median	quant75	quant90	max
1 66	0.2261038	0.3916233	0.3916233	0.4090372	0.3916233	0.4522076	0.4522076	0.4522076	

False positives resulting from the use of various LSD statistics

	c	min	quant10	quant25	mean	median	quant75	quant90	max
false.pos 66	7	2	2	2	2	0	0	0	

False negatives resulting from the use of various LSD statistics

	c	min	quant10	quant25	mean	median	quant75	quant90	max
false.neg 66	0	0	0	2	0	4	4	4	

The function `findLSDminerrors` can be used to find a value of the LSD that minimizes the number of false positive (negative) errors i.e. declaring a pairwise difference to be significant (not significant) when it is not significant (significant). The function has an argument `false.pos.wt` that specifies how many false negatives are equivalent to one false positive; the default for `false.pos.wt` is 10, which is a conservative approach because it greatly favours false negatives over false positives.

```
(findLSDminerrors(diffs))
```

	LSD	false.pos	false.neg	false.criterion
overall	0.4489466	0	4	4

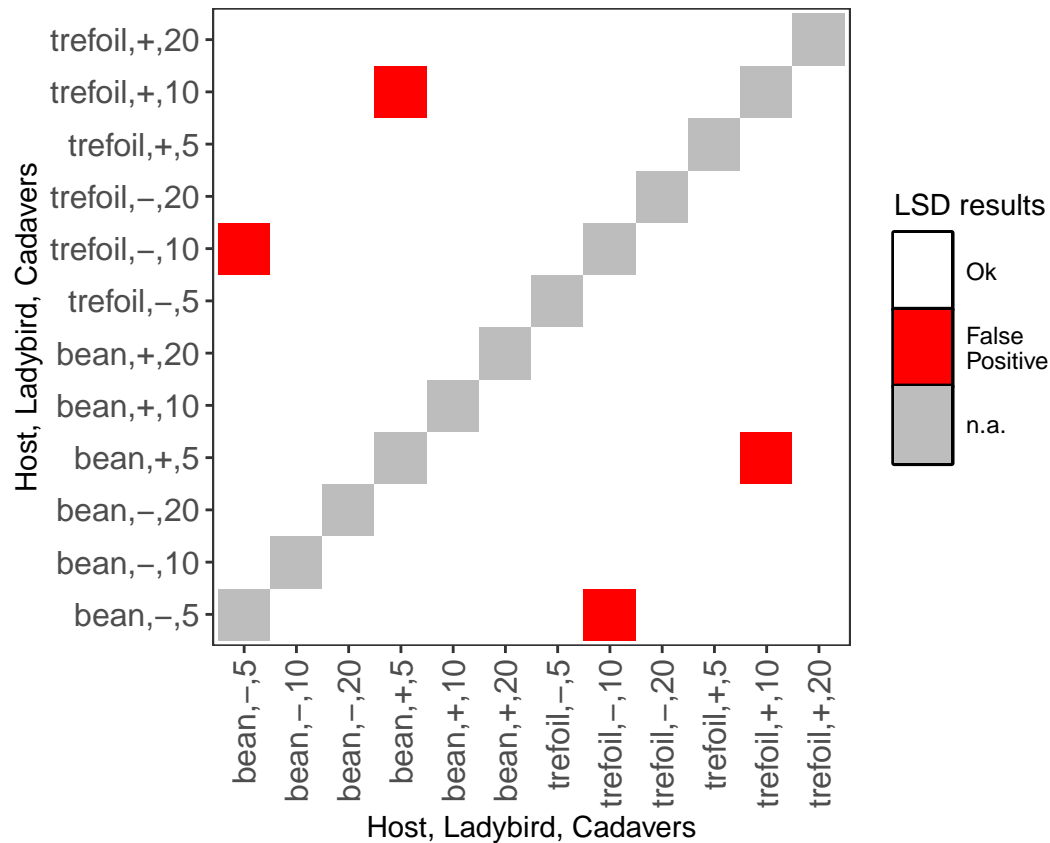
```
(minLSDpos <- findLSDminerrors(diffs, false.pos.wt = 1))
```

	LSD	false.pos	false.neg	false.criterion
overall	0.354983	2	0	2

The smallest value of the LSD that results in the minimum weighted sum of the false positives and negatives is 0.4489466 when the weight is 10 and is 0.354983 when the weight is one. These values result in 4 false negatives and 2 false positives respectively.

To see which of the two pairwise comparisons will be falsely identified as being significant when the LSD value is set to minimum LSD, the LSDs stored in the `alldiffs` object need to be recalculated to be based on this LSD value. Then plot the errors or save the return values obtained using the function `plotLSDerrors`. The plot below shows that the two pairs whose differences are incorrectly identified as significant have the same level of the Ladybird factor, but differ in both of the levels for the Host and Cadaver factors.

```
diffs.overall <- recalcLSD(diffs, LSDtype = "supplied", LSDsupplied = minLSDpos["LSD"])
plotLSDerrors(diffs.overall)
```



This raises the question of whether the minimum LSD should be used. There are at least four alternatives: (i) use it without restriction, on the basis that it can be concluded that using it is unlikely to result in seriously flawed conclusions; (ii) use it with the restriction that it only be applied to assess pairwise comparisons that have the same Host or the same Cadaver treatment; (iii) investigate the use of the minimum LSD with only false negative errors; and (iv) rather than use an overall LSD value, use LSD values computed from the LSDs within each Host level.

Because `LSDtype` was set to `"factor.combination"` and `LSDby` to `"Host"` in forming the object `diffs`, the LSDs for alternative (iv) are stored in the LSD component of the object `diffs`. Printing out the LSD component will summarize how those LSD values perform. Otherwise, the following call to `exploreLSDs` will display the properties of the LSDs for various LSD statistics:

```
exploreLSDs(diffs, LSDtype = "factor.combination", LSDby = "Host")
```

The following shows the contents of the LSD component of `diffs`:

```
(diffs$LSD)
```

	c	minLSD	meanLSD	maxLSD	assignedLSD	accuracyLSD	falsePos	falseNeg
bean	15	0.3916233	0.3916233	0.3916233	0.3916233	8.504778e-16	0	0
trefoil	15	0.3916233	0.3916233	0.3916233	0.3916233	7.087315e-16	0	0

Because the minimum and maximum LSD values are equal, it follows that there is only one value of the LSD for all pairwise comparisons within each Host level and it happens that the values for the two Hosts are also equal. The table shows that zero false positive and negative errors will result from the use of the value of 0.39 for the 30 within-Host comparisons. Thus, using `findLSDminerrors(diffs, LSDtype = "factor.combinations", LSDby = "Host")` to select the minimum LSDs would only confirm what is already obvious.

Plot the predictions

```
titl <- str2expression("logit~(italic(P))")
names(titl) <- "logitP"
plotPredictions(diffs$predictions, y = "predicted.value",
  y.title = titl,
  classify = "Host:Ladybird:Cadavers",
  error.intervals = "halfLeast", interval.annotate = FALSE,
  ggplotFuncs = list(facet_grid(Ladybird ~ Host,
    labeller = label_both)))
```

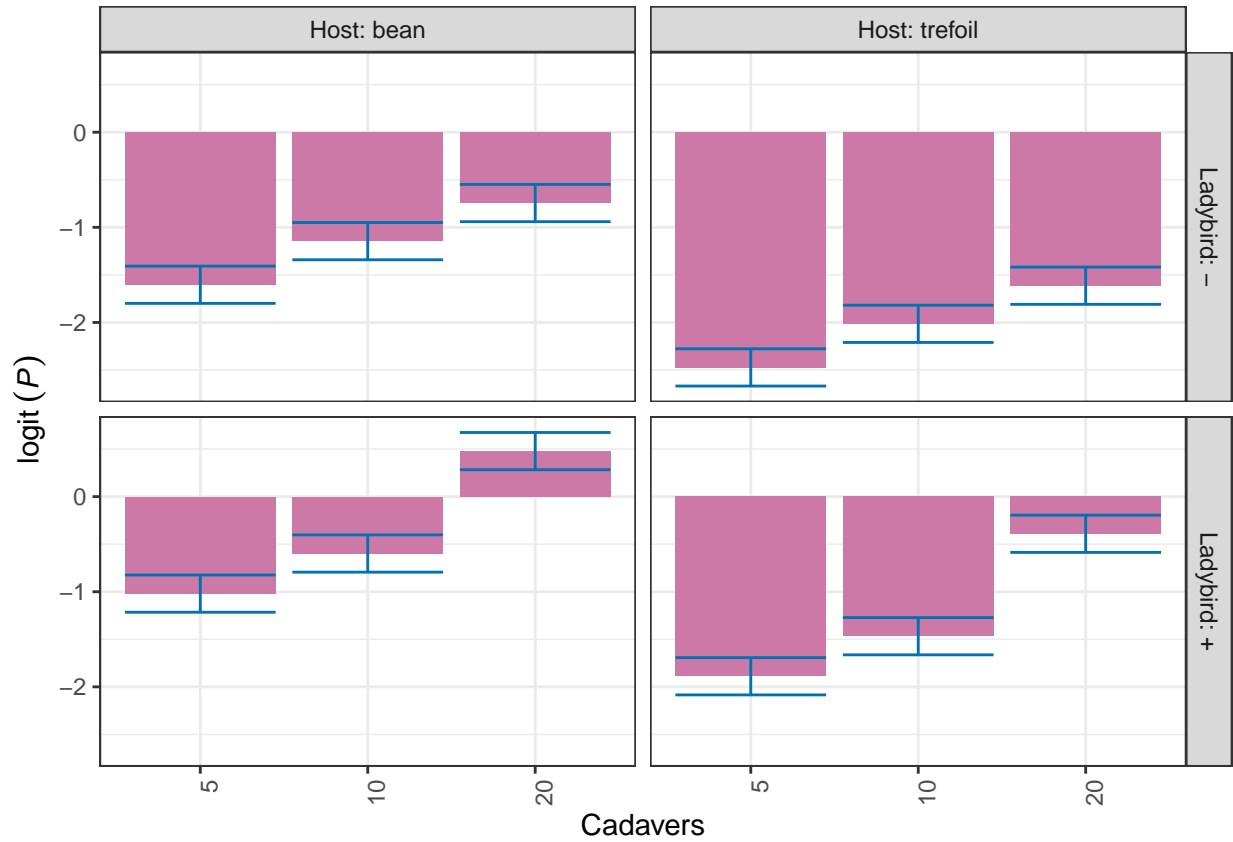


Figure 1. Estimated marginal means (EMMs) for $\text{logit}(P)$, where P is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels. Error bars are an EMM \pm half-LSD (5%). The two EMMs for the same Host are significantly different ($p \leq 0.05$) if their error bars do not overlap.

The function `plotPredictions` uses `ggplot` to produce the plot and the `ggplotFuncs` argument allows the addition of `ggplot` functions to modify the plot. In this case, the `facet.grid` function is respecified to include `prepender` functions that modify the labels of the facets to include the factor names. Note the statement in the legend of Figure 1 that restricts the use of the error bars to determining the significance of differences for the pairwise comparisons of EMMs for the same Host.

Get and plot the predictions with a single function call

The `predictPresent` function combines the functionality of `predictPlus` and `plotPredictions`, as demonstrated now. Also, the use of `plotPvalues` to plot the pairwise p -values is displayed. The `predictPresent` function has the capability of producing `alldiffs` objects for multiple terms and these are stored in a list each of which is named for the term whose `alldiffs` object it stores. Thus, the term has to be specified in referencing components of `diffs`.

```
titl <- "logit(P)"
names(titl) <- "logitP"
diffs <- predictPresent(current.asrt$asreml.obj,
                        terms = "Host:Ladybird:Cadavers",
                        linear.transformation = ~Cadavers:Ladybird + Host,
                        titles = titl,
                        wald.tab = current.asrt$wald.tab,
                        error.intervals = "halfLeast",
                        LSDtype = "factor.combination", LSDby = "Host",
                        tables = "none",
                        ggplotFuncs = list(facet_grid(Ladybird ~ Host,
                                                    labeller = label_both)))
```

Joining with 'by = join_by(fac.comb)'

Joining with 'by = join_by(Host)'

```
plotPvalues(diffs$Host.Ladybird.Cadavers, factors.per.grid = 1, show.sig = TRUE)
```

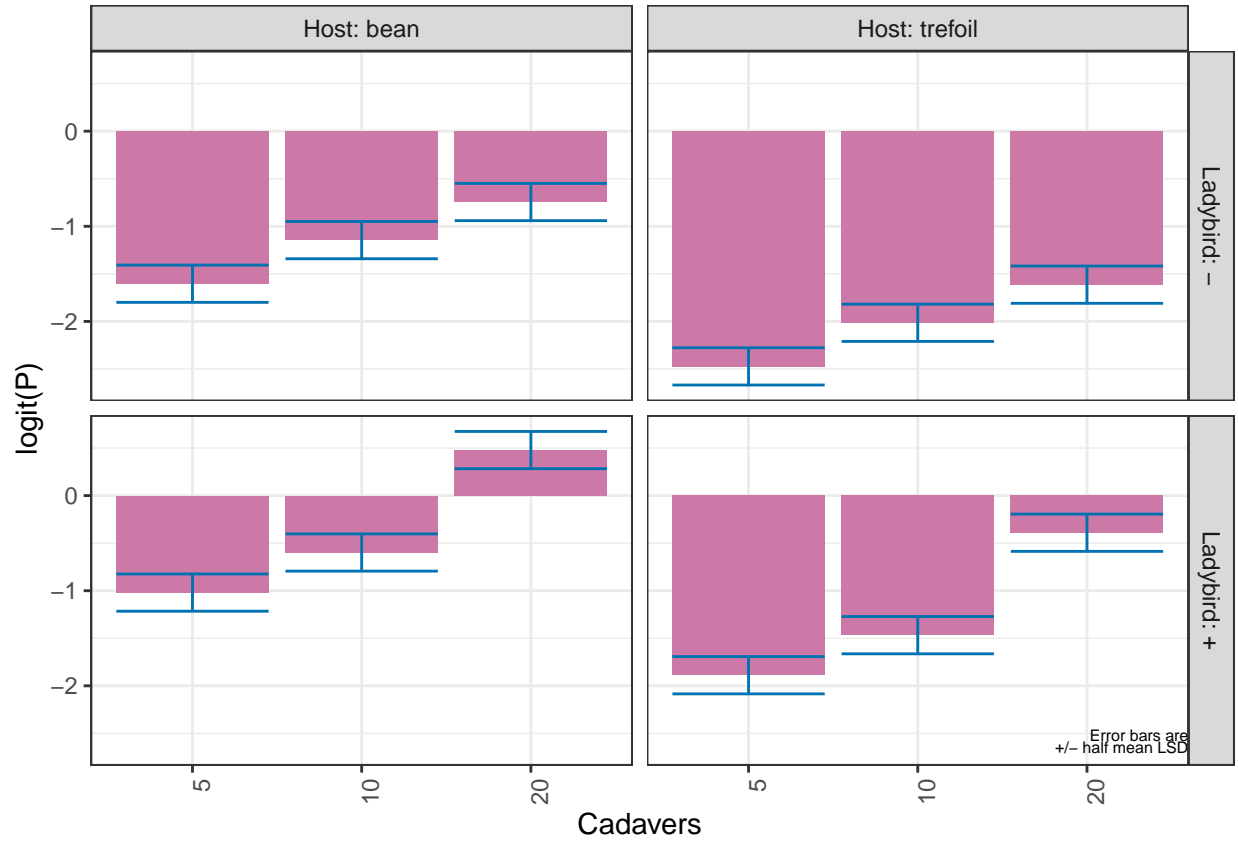


Figure 2. Estimated marginal means (EMMs) for $\text{logit}(P)$, where P is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels. Error bars are an EMM \pm half-LSD (5%). The two EMMs for the same Host are significantly different ($p \leq 0.05$) if their error bars do not overlap.

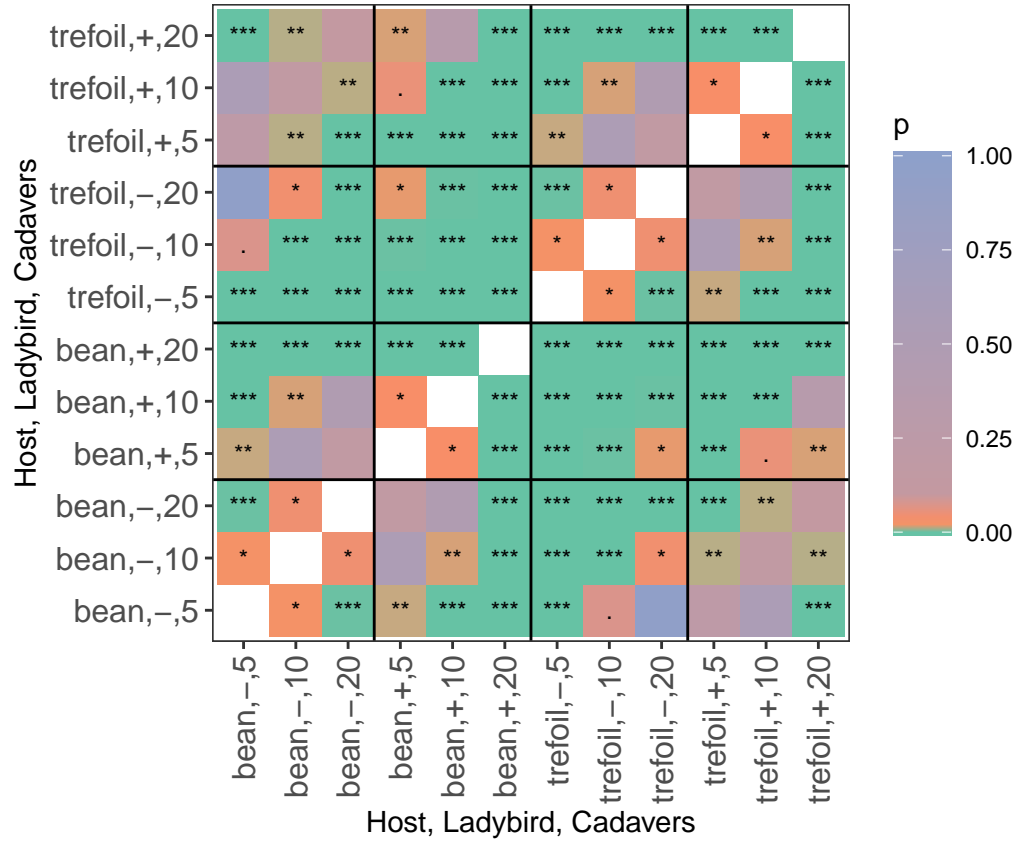


Figure 3. The p -values for each of the pairwise comparisons of the estimated marginal means for $\text{logit}(P)$, where P is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels

```
options(width = 90)
diffs$Host.Ladybird.Cadavers$differences
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20
bean,-,5	0.00000000	-0.4584030	-0.8590241	-0.5842863	-1.0054898	-2.0825042
bean,-,10	0.45840297	0.00000000	-0.4006211	-0.1258833	-0.5470869	-1.6241012
bean,-,20	0.85902408	0.4006211	0.00000000	0.2747378	-0.1464657	-1.2234801
bean,+,5	0.58428627	0.1258833	-0.2747378	0.00000000	-0.4212036	-1.4982179
bean,+,10	1.00548982	0.5470869	0.1464657	0.4212036	0.00000000	-1.0770144
bean,+,20	2.08250420	1.6241012	1.2234801	1.4982179	1.0770144	0.00000000
trefoil,-,5	-0.86920012	-1.3276031	-1.7282242	-1.4534864	-1.8746899	-2.9517043
trefoil,-,10	-0.41079715	-0.8692001	-1.2698212	-0.9950834	-1.4162870	-2.4933014
trefoil,-,20	-0.01017604	-0.4685790	-0.8692001	-0.5944623	-1.0156659	-2.0926802
trefoil,+,5	-0.28491385	-0.7433168	-1.1439379	-0.8692001	-1.2904037	-2.3674180
trefoil,+,10	0.13628970	-0.3221133	-0.7227344	-0.4479966	-0.8692001	-1.9462145
trefoil,+,20	1.21330408	0.7549011	0.3542800	0.6290178	0.2078143	-0.8692001
	trefoil,-,5	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20
bean,-,5	0.8692001	0.4107972	0.01017604	0.2849139	-0.1362897	-1.2133041
bean,-,10	1.3276031	0.8692001	0.46857901	0.7433168	0.3221133	-0.7549011
bean,-,20	1.7282242	1.2698212	0.86920012	1.1439379	0.7227344	-0.3542800
bean,+,5	1.4534864	0.9950834	0.59446231	0.8692001	0.4479966	-0.6290178
bean,+,10	1.8746899	1.4162870	1.01566586	1.2904037	0.8692001	-0.2078143
bean,+,20	2.9517043	2.4933014	2.09268024	2.3674180	1.9462145	0.8692001
trefoil,-,5	0.00000000	-0.4584030	-0.85902408	-0.5842863	-1.0054898	-2.0825042
trefoil,-,10	0.4584030	0.00000000	-0.40062111	-0.1258833	-0.5470869	-1.6241012
trefoil,-,20	0.8590241	0.4006211	0.00000000	0.2747378	-0.1464657	-1.2234801
trefoil,+,5	0.5842863	0.1258833	-0.27473781	0.00000000	-0.4212036	-1.4982179
trefoil,+,10	1.0054898	0.5470869	0.14646574	0.4212036	0.00000000	-1.0770144
trefoil,+,20	2.0825042	1.6241012	1.22348012	1.4982179	1.0770144	0.00000000

```
options(width = 90)
print(diffs$Host.Ladybird.Cadavers$sed)
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20	trefoil,-,5
bean,-,5	NA	0.1957142	0.1957142	0.1957142	0.1957142	0.1957142	0.1129957
bean,-,10	0.1957142	NA	0.1957142	0.1957142	0.1957142	0.1957142	0.2259913
bean,-,20	0.1957142	0.1957142	NA	0.1957142	0.1957142	0.1957142	0.2259913
bean,+,5	0.1957142	0.1957142	0.1957142	NA	0.1957142	0.1957142	0.2259913
bean,+,10	0.1957142	0.1957142	0.1957142	0.1957142	NA	0.1957142	0.2259913
bean,+,20	0.1957142	0.1957142	0.1957142	0.1957142	0.1957142	NA	0.2259913
trefoil,-,5	0.1129957	0.2259913	0.2259913	0.2259913	0.2259913	0.2259913	NA
trefoil,-,10	0.2259913	0.1129957	0.2259913	0.2259913	0.2259913	0.2259913	0.1957142
trefoil,-,20	0.2259913	0.2259913	0.1129957	0.2259913	0.2259913	0.2259913	0.1957142
trefoil,+,5	0.2259913	0.2259913	0.2259913	0.1129957	0.2259913	0.2259913	0.1957142
trefoil,+,10	0.2259913	0.2259913	0.2259913	0.2259913	0.1129957	0.2259913	0.1957142
trefoil,+,20	0.2259913	0.2259913	0.2259913	0.2259913	0.2259913	0.1129957	0.1957142
	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20		
bean,-,5	0.2259913	0.2259913	0.2259913	0.2259913	0.2259913		
bean,-,10	0.1129957	0.2259913	0.2259913	0.2259913	0.2259913		
bean,-,20	0.2259913	0.1129957	0.2259913	0.2259913	0.2259913		
bean,+,5	0.2259913	0.2259913	0.1129957	0.2259913	0.2259913		
bean,+,10	0.2259913	0.2259913	0.2259913	0.1129957	0.2259913		
bean,+,20	0.2259913	0.2259913	0.2259913	0.2259913	0.1129957		
trefoil,-,5	0.1957142	0.1957142	0.1957142	0.1957142	0.1957142		
trefoil,-,10	NA	0.1957142	0.1957142	0.1957142	0.1957142		
trefoil,-,20	0.1957142	NA	0.1957142	0.1957142	0.1957142		
trefoil,+,5	0.1957142	0.1957142	NA	0.1957142	0.1957142		
trefoil,+,10	0.1957142	0.1957142	0.1957142	NA	0.1957142		
trefoil,+,20	0.1957142	0.1957142	0.1957142	0.1957142	NA		

Perform the analysis with just selected model fitted

The model with nonsignificant fixed terms dropped is obtained in order to compare it with the fit when they are retained and the estimated marginal means for the chosen model are obtained.

```
ns.terms <- current.asrt$test.summary$terms[current.asrt$test.summary$action == "Nonsignificant"]
red.asrt <- changeTerms(current.asrt, dropFixed = paste(ns.terms, collapse = "+"))
```

```
WARN [2025-06-10 18:08:12] Some components changed by more than 1% on the last iteration
```

```
Warning in asreml(fixed = logitP ~ Host + Cadavers + Ladybird + Cadavers:Ladybird, : Some
components changed by more than 1% on the last iteration
```

```
WARN [2025-06-10 18:08:12] Some components changed by more than 1% on the last iteration
```

```
Warning in asreml(fixed = logitP ~ Host + Cadavers + Ladybird + Cadavers:Ladybird, : Some
components changed by more than 1% on the last iteration
```

```
summary(red.asrt$asreml.obj)$varcomp
```

	component	std.error	z.ratio	bound	%ch
Run	-0.004328175	0.002777746	-1.558161	U	1.1
Run:Plant!R	0.223456033	0.039510023	5.655680	P	0.0

```
print(red.asrt, which = "pseudoanova")
```

```
#### Pseudo-anova table for fixed terms
```

```
Wald tests for fixed effects.
```

```
Response: logitP
```

	Df	denDF	F.inc	Pr
(Intercept)	1	1	1551.00	0.0162
Host	1	64	60.88	0.0000
Cadavers	2	64	38.12	0.0000
Ladybird	1	64	49.65	0.0000
Cadavers:Ladybird	2	64	3.88	0.0256

```
diffs.red <- predictPlus(red.asrt$asreml.obj,
  classify = "Host:Ladybird:Cadavers",
  wald.tab = current.asrt$wald.tab,
  error.intervals = "halfLeast", interval.annotate = FALSE,
  LSDtype = "factor.combination", LSDby = "Host",
  tables = "predictions")
```

```
Joining with 'by = join_by(fac.comb)'
```

```
Joining with 'by = join_by(Host)'
```

Predictions for logitP from Host:Ladybird:Cadavers

Notes:

- The predictions are obtained by averaging across the hypertable calculated from model terms constructed solely from factors in the averaging and classify sets.
- Use 'average' to move ignored factors into the averaging set.
- The ignored set: Run

	Host	Ladybird	Cadavers	predicted.value	standard.error
1	bean	-	5	-1.6038338	0.1398332
2	bean	-	10	-1.1454308	0.1398332
3	bean	-	20	-0.7448097	0.1398332
4	bean	+	5	-1.0195475	0.1398332
5	bean	+	10	-0.5983440	0.1398332
6	bean	+	20	0.4786704	0.1398332
7	trefoil	-	5	-2.4730339	0.1398332
8	trefoil	-	10	-2.0146309	0.1398332
9	trefoil	-	20	-1.6140098	0.1398332
10	trefoil	+	5	-1.8887476	0.1398332
11	trefoil	+	10	-1.4675441	0.1398332
12	trefoil	+	20	-0.3905297	0.1398332
	upper.halfLeastSignificant.limit			lower.halfLeastSignificant.limit	est.status
1	-1.4107942			-1.7968734	Estimable
2	-0.9523913			-1.3384704	Estimable
3	-0.5517702			-0.9378493	Estimable
4	-0.8265080			-1.2125871	Estimable
5	-0.4053044			-0.7913835	Estimable
6	0.6717100			0.2856308	Estimable
7	-2.2799944			-2.6660735	Estimable
8	-1.8215914			-2.2076705	Estimable
9	-1.4209703			-1.8070494	Estimable
10	-1.6957081			-2.0817872	Estimable
11	-1.2745045			-1.6605837	Estimable
12	-0.1974902			-0.5835693	Estimable

LSD values

minimum LSD = 0.3860791 0.3860791

mean LSD = 0.3860791 0.3860791

maximum LSD = 0.3860791 0.3860791

(sed range / mean sed = 1.29e-15 1.29e-15)

```
options(width = 90)
print(diffs.red$sed)
```

bean,-,5 bean,-,10 bean,-,20 bean,+,5 bean,+,10 bean,+,20 trefoil,-,5

bean,-,5	NA	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	0.1113960
bean,-,10	0.1929435	NA	0.1929435	0.1929435	0.1929435	0.1929435	0.2227920
bean,-,20	0.1929435	0.1929435	NA	0.1929435	0.1929435	0.1929435	0.2227920
bean,+,5	0.1929435	0.1929435	0.1929435	NA	0.1929435	0.1929435	0.2227920
bean,+,10	0.1929435	0.1929435	0.1929435	0.1929435	NA	0.1929435	0.2227920
bean,+,20	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	NA	0.2227920
trefoil,-,5	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	NA
trefoil,-,10	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920	0.1929435
trefoil,-,20	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920	0.1929435
trefoil,+,5	0.2227920	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920	0.1929435
trefoil,+,10	0.2227920	0.2227920	0.2227920	0.2227920	0.1113960	0.2227920	0.1929435
trefoil,+,20	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	0.1113960	0.1929435
	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20		
bean,-,5	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920		
bean,-,10	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920		
bean,-,20	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920		
bean,+,5	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920		
bean,+,10	0.2227920	0.2227920	0.2227920	0.1113960	0.2227920		
bean,+,20	0.2227920	0.2227920	0.2227920	0.2227920	0.1113960		
trefoil,-,5	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435		
trefoil,-,10	NA	0.1929435	0.1929435	0.1929435	0.1929435		
trefoil,-,20	0.1929435	NA	0.1929435	0.1929435	0.1929435		
trefoil,+,5	0.1929435	0.1929435	NA	0.1929435	0.1929435		
trefoil,+,10	0.1929435	0.1929435	0.1929435	NA	0.1929435		
trefoil,+,20	0.1929435	0.1929435	0.1929435	0.1929435	NA		

References

- Brien, C. J. (2025a) *asremlPlus: Augments ASReml-R in fitting mixed models and packages generally in exploring prediction differences*. Version 4.4.49. <https://cran.r-project.org/package=asremlPlus/> or <http://chris.brien.name/rpackages/>.
- Brien, C. J. (2024b) *dae: Functions useful in the design and ANOVA of experiments*. Version 3.2.30. <https://cran.r-project.org/package=dae/> or <http://chris.brien.name/rpackages/>.
- Butler, D. G., Cullis, B. R., Gilmour, A. R., Gogel, B. J. and Thompson, R. (2023). *ASReml-R Reference Manual Version 4.2*. VSN International Ltd, <https://asreml.kb.vsnr.co.uk/>.
- Littell, R. C., Milliken, G. A., Stroup, W. W., Wolfinger, R. D., & Schabenberger, O. (2006). *SAS for Mixed Model.* (2nd ed.). Cary, N.C.: SAS Press.
- R Core Team (2025) *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.r-project.org/>.
- Snee, R. D. (1981). Graphical Display and Assessment of Means. *Biometrics*, **37**, 835–836.
- Welham, S. J., Gezan, S. A., Clark, S. J., & Mead, A. (2014). *Statistical Methods in Biology: Design and Analysis of Experiments and Regression*. Boca Raton: Chapman and Hall/CRC.